Resistance to Root-knot Nematodes in Cucumber and Horned Cucumber

TODD C. WEHNER, S. ALAN WALTERS, AND KENNETH R. BARKER²

Abstract: Two experiments were conducted in the greenhouse. In one experiment, cucumber (Cucumis sativus) and horned cucumber (C. metuliferus) cultigens were evaluated for resistance to four root-knot nematode species (Meloidogyne arenaria, M. hapla, M. incognita, and M. javanica), and, in a second experiment, a standard (12-week) test was compared with a rapid (6-week) test. In the first experiment, horned cucumber cultigens varied in response to the Meloidogyne species. 'Sumter' cucumber was more susceptible than the horned cucumber to Meloidogyne incognita, M. javanica, and M. arenaria. All cultigens were more resistant to M. hapla than to the other root-knot nematode species. In the second experiment, best results were obtained when the test was run for 12 weeks rather than 6 weeks after planting (or 10 and 4 weeks after inoculation, respectively). All cultigens were more resistant to M. arenaria than to either M. incognita or M. javanica.

Key words: African horned cucumber, cucumber, Cucumis metuliferus, Cucumis sativus, Meloidogyne arenaria, Meloidogyne hapla, Meloidogyne incognita, Meloidogyne javanica, nematode resistance, root-

knot nematode.

Cucumber (Cucumis sativus L.) is the second most important vegetable crop in North Carolina, and root knot (caused by Meloidogyne spp.) is the most important cucumber disease. Annually, root-knot nematodes destroy approximately 12% of the crop in the state (6). Four species of root-knot nematodes can potentially attack cucumber in North Carolina: Meloidogyne incognita (Kofoid & White) Chitwood, M. arenaria (Neal) Chitwood, M. hapla Chitwood, and M. javanica (Treub) Chitwood; but M. hapla is not important on cucumber in the state. Cucumber cultivars resistant to these species would be valuable for growers.

The African horned cucumber (Cucumis metuliferus Naud.) is resistant but not immune to M. arenaria, M. javanica, and M. incognita (3). As a result, interspecific hybridization to transfer root-knot nematode resistance to the cultivated cucumber has been a goal of many researchers. So far, attempts to hybridize the two species have been unsuccessful (4).

Screening methods have been standardized (2,7) for the accurate testing of cultigens (used to refer to breeding lines, cultivars, and plant introduction accessions collectively) for resistance to root-knot nematodes. This research involved two greenhouse experiments, one conducted in 1987 and one in 1988. The objective of the first experiment was to determine whether there were differences among C. metuliferus cultigens for resistance to four root-knot nematode species. The objective of the second experiment was to determine whether a 6-week test for rapid screening in a breeding program would predict the results of the standard 12-week test in determining the resistance of Cucumis cultigens to root-knot nematodes.

MATERIALS AND METHODS

Nematode species experiment: During the spring of 1987, five accessions of C. metu-liferus and 'Sumter' cucumber (C. sativus) were evaluated for resistance to four species of root-knot nematodes (M. incognita race 1, M. javanica, M. arenaria race 1, and M. hapla). The experiment was a factorial treatment arrangement in a randomized complete block design with four replications. Each treatment combination consisted of one Cucumis cultigen and one nematode species in a 100-mm diameter

Received for publication 21 February 1991.

* Professor, Department of Plant Pathology, North Carolina State University, Raleigh, NC 27695-7616.

¹ Professor and graduate student, Department of Horticultural Science, North Carolina State University, Raleigh, NC 27695-7609.

The authors gratefully acknowledge the assistance of Mr. Rufus R. Horton, Jr., Mr. Kerrick M. Hartman, and Dr. Joseph N. Sasser.

(1,450 cm³ volume) pot. Each pot contained a 1:1 mix of steam-sterilized sand and field soil (loamy sand: 80% sand, 15% silt, and 5% clay). Five seeds were planted directly into each pot. Pots were irrigated twice daily using drip irrigation with fertilizer injection. At the three-leaf stage, plants were thinned to one per pot. Greenhouse temperature was 24–32 C day and 21–24 C night.

Each plant was inoculated 2 weeks after planting with 5,000 nematode eggs (5). Plants were evaluated for root-knot nematode galling 8 weeks after inoculation (10 weeks after planting) by rating the percentage (0 to 100%) of the root system galled (1). The number of egg masses per root system was also counted. Finally, roots were cut into segments 10 to 20 mm long and treated with 1% NaOCl (5) to extract the eggs from the root system. Gall index, egg number, and egg mass number were analyzed with the GLM procedure of SAS (SAS Institute, Cary, NC). Cultigen means for gall index, egg number, and egg mass number were compared using Fisher's LSD at P = 0.05.

Six-week vs. 12-week experiment: In spring 1988, an experiment was conducted to determine whether a rapid (6-week) test was as good as the standard (12-week) test for evaluating resistance to root-knot nematode. Resistance of four C. sativus and two C. metuliferus cultigens to M. incognita race 1, M. incognita race 3, M. arenaria race 1, M. arenaria race 2, and M. javanica was determined with the two test methods.

The experiment was a factorial treatment arrangement in a randomized complete block design with three replications. The treatment combination consisted of one plant, one method, and one root-knot nematode species or race per pot. Each pot contained a 1:1 mix of steam-sterilized sand and field soil (loamy sand: 80% sand, 15% silt, and 5% clay). Five seeds were sown into 100-mm diameter, 1,450 cm³ pots, which were irrigated twice daily by drip irrigation with fertilizer injection.

At the two-leaf stage, one plant was taken out of each pot and transplanted into larger (150-mm diameter, 1700 cm³) pots. At the three-leaf stage, the remaining plants were thinned to one per small pot. The plants in the small pots were used for the 6-week test; those in large pots for the 12week test. Each treatment combination was inoculated 2 weeks after planting with 5,000 nematode eggs (5). Greenhouse temperature was 24–32 C day and 21–24 C night.

Each treatment combination was evaluated 6 or 12 weeks after planting (corresponding to 4 and 10 weeks after inoculation), depending on test method. Plants were evaluated by rating the roots for percentage galled by root-knot nematodes (1). Gall index data were analyzed with the GLM procedure of SAS. Means for cultigen and method were separated using Fisher's LSD at P = 0.05.

RESULTS AND DISCUSSION

Nematode species experiment: For our purposes, the percentage of the root system galled can be used to classify resistance: 0-10% = highly resistant; 11-20% = moderately resistant; 21-40% = slightly resistant; and > 40% = susceptible. By these criteria, all Cucumis cultigens evaluated had some resistance to M. hapla (Table 1). 'Sumter' cucumber was susceptible to all Meloidogyne spp., except for M. hapla. The cultigens of C. metuliferus were susceptible to both M. incognita race 1 and M. javanica. Gall indices among C. metuliferus cultigens differed in response to two root-knot nematodes, M. arenaria race 1 and M. incognita race 1. Galling from M. arenaria race 1 and M. javanica was lower (P < 0.05) on all C. metuliferus cultigens than on C. sativus cv. Sumter. Over all treatment combinations, gall index and egg number were highly correlated (r = 0.79).

Numbers of eggs and egg masses of M. incognita race 1, M. arenaria race 1, and M. javanica were greater ($P \le 0.05$) on 'Sumter' cucumber than on the C. metaliferus cultigens. For M. incognita race 1, PI 482452 had fewer ($P \le 0.05$) eggs than the other C. metaliferus cultigens, indicating a higher level of resistance. No significant differ-

TABLE 1. Gall index (GI)†, number of eggs, and number of egg masses (EM) on six cultigens of Cucumis spp. tested with four species of root-knot nematodes (Meloidogyne spp.).

		M. hapla			M. arenaria			M. incognita			M. javanica		
Cultigen	Species	GI	Eggs	EM	GI	Eggs	EM	GI	Eggs	EM	GI	Eggs	EM
PI 482452	C. metuliferus	6	220	0	36	700	0	50	2,550	1	46	3,605	1
PI 482454	C. metuliferus	6	5	0	40	1,785	0	60	22,290	2	60	6,440	2
PI 482450	C. metuliferus	6	220	0	60	2,747	1	66	21,547	1	41	2,167	0
PI 482461	C. metuliferus	16	305	0	56	1.015	1	80	26,390	2	46	4.820	1
PI 482448	C. metuliferus	20	165	0	50	3,420	1	86	22,400	3	66	7.250	1
Sumter	C. sativus	30	255	0	96	69,200	5	96	63,000	4	96	75,000	5
LSD $(P = 0.09)$	5) for row and colu	mn co	mpari	sons:	22	12,639	1		263/180000000			A. (1987) 150	

Data are means of four replications of one plant each. Meloidogyne arenaria and M. incognita were race 1. Plants were inoculated 2 weeks after planting and evaluated 8 weeks later.

† Gall index = percentage of the root system galled (1).

ences ($P \le 0.05$) occurred among the *Cucumis* cultigens for number of eggs or egg masses of *M. hapla*, indicating that all were resistant. Based on gall indices, number of eggs, and number of egg masses, PI 482452 was the most resistant cultigen to the root-knot nematodes tested. Although counts of eggs are preferable to galling indices for obtaining an indication of nematode reproduction (i.e., resistance), the galling indices may be advantageous when large

numbers of cultigens must be assessed and time or labor is limited.

Six-week vs. 12-week experiment: The 6-week test did not allow time for extensive nematode gall development on susceptible plants. In some cases, it also did not allow for differentiation between C. metalliferus and C. sativus cultigens (Table 2). In the 12-week test, more of the differences between the two species were significant ($P \le 0.05$).

Cucumis spp. were more resistant to races

TABLE 2. Gall index† for six cultigens of Cucumis spp. tested with five species of root-knot nematodes (Meloidogyne spp.) and two methods.

		Gall indices						
Cultigen	Species	Mi1	Mi3	Mj	Mal	Ma2		
	Six-	week test						
PI 482452	C. metuliferus	21	29	19	14	10		
PI 482448	C. metuliferus	18	28	18	17	12		
Wisconsin SMR 18	C. sativus	31	53	24	17	11		
Poinsett 76	C. sativus	32	54	36	19	12		
Marketmore 76	C. sativus	40	57	40	25	16		
Sumter	C. sativus	25	52	32	20	11		
	Twelv	e-week test						
PI 482452	C. metuliferus	24	16	10	10	10		
PI 482448	C. metuliferus	26	16	24	13	9		
Wisconsin SMR 18	C. sativus	95	89	93	15	14		
Poinsett 76	C. sativus	95	95	95	24	18		
Marketmore 76	C. sativus	96	95	97	26	17		
Sumter	C. sativus	98	99	98	44	26		
Mean over cultigens and	50	57	49	20	14			
LSD $(P = 0.05)$ for row			10					

Data are means of three replications of one plant each. The 6-week test used small (100-mm diameter) pots for 6 weeks (planting to evaluation), and the 12-week test used large (150-mm diameter) pots for 12 weeks (planting to evaluation).

† Gall index = percentage of the root system galled (1). Mi1 = M. incognita race 1, Mi3 = M. incognita race 3, Mj = M. javanica, Ma1 = M. arenaria race 1, and Ma2 = M. arenaria race 2.

of M. arenaria than to either M. incognita or M. javanica. Across all cultivars in both 6-week and 12-week tests, mean gall indices were lower for M. arenaria races 1 and 2 than for M. incognita races 1 and 3 or M. javanica. We concluded that the 12-week test was worth the extra time, because the 6-week test failed to detect important differences. The 6-week test (4 weeks after inoculation) does not allow sufficient time for root-knot nematode reproduction.

Two check lines that are often used in screening for resistance to foliar diseases in cucumbers are Wisconsin SMR 18 and Sumter. In the 12-week test, Wisconsin SMR 18 was moderately resistant (15% galling) to M. arenaria, whereas Sumter (35% galling) was only slightly resistant to M. arenaria (figures averaged across races 1 and 2). Sumter cucumber was highly susceptible to M. arenaria race 1 in the nematode species test but was moderately resistant to the same root-knot nematode race in the 6-week vs. 12-week experiment. This may have occurred because different populations were used in the two experiments.

Accessions of *C. metuliferus* were not highly resistant to the root-knot nematode species, *M. arenaria*, *M. incognita*, and *M. javanica*. However, there was some resistance in some accessions, agreeing with the findings of Dalmasso et al. (3). Specific races of *M. incognita* and *M. arenaria* were not described by Dalmasso et al. (3), only that

three populations of each were tested. Our studies have provided additional information on *M. hapla* and on specific races of *M. arenaria* and *M. incognita*. Additional research is needed, particularly to evaluate the *Cucumis* germplasm collection for nematode resistance by the 12-week test method.

LITERATURE CITED

 Barker, K. R. 1985. Nematode extraction and bioassays. Pp. 19-35 in K. R. Barker, C. C. Carter, and J. N. Sasser, eds. An advanced treatise on Meloidogyne, vol. 2: Methodology. Raleigh: North Carolina State University Graphics.

 Boukema, I. W., G. T. M. Reuling, and K. Hofman. 1984. The reliability of a seedling test for resistance to root-knot nematodes in Cucurbits. Cucurbit Genetics Cooperative Report 7:92–93.

 Dalmasso, A., R. D. de Vaulx, and M. Pitrat. 1981. Responses of some Cucurbita and Cucumis accessions to three Meloidogyne species. Cucurbit Genetics Gooperative Report 4:53-55.

 Deakin, J. R., G. W. Bohn, and T. W. Whitaker. 1971. Interspecific hybridization in *Gucumis*. Economic Botany 25:195–211.

 Hussey, R. S., and K. R. Barker. 1973. A comparison of methods of collecting inocula for *Meloi-dogyne* spp., including a new technique. Plant Disease Reporter 57:1025–1028.

 Main, C. E., and S. K. Gurtz, editors. 1989.
 1988 estimates of crop losses in North Carolina due to plant diseases and nematodes. Department of Plant Pathology Special Publication No. 8, North Carolina State University, Raleigh.

Nijs, A. P. M. den, and K. Hofman. 1983. An
efficient procedure to screen for resistance to rootknot nematodes in cucurbits. Cucurbit Genetics Cooperative Report 6:96–98.